



eISSN 2330-0280

VEDIC RESEARCH INTERNATIONAL

PHYTOMEDICINE

JOURNAL HOME PAGE AT WWW.VEDICJOURNALS.COM

REVIEW

DOI: <http://dx.doi.org/10.14259/pm.v1i2.39>

Current Status of Indian Medicinal Plants with Immunomodulatory Potential: A Review

KALLIAPPAN ILANGO^{1*}, ANANTH KUMAR KAMMALLA¹, GOVIND PRASAD DUBEY², ARUNA AGRAWAL³

¹Interdisciplinary School of Indian System of Medicine, SRM University, Kattankulathur-603203, Tamilnadu, India.

²National Facility for Tribal & Herbal Medicine, Institute of Medical sciences, ³Faculty of Ayurveda, Institute of Medical sciences, Banaras Hindu University, Varanasi, India.

Article Info: Received: June 28th, 2013; Accepted: June 29th, 2013

ABSTRACT

Every country in the world has lists of herbal remedies for the treatment of diseases and different other unwanted conditions in humans. Ayurveda is one of the traditional systems of medicine practiced in India and Sri Lanka and can be traced back to 6000 B.C. Modulation of immune responses to alleviate the diseases has been of interest for many years and the concept of 'Rasayana' in Ayurveda is based on related principles. Rasayana is helpful to improve immunity and is normally advised during the degenerative phase of life, which starts from around 45 years in both men and women. Some of the medicinal plants valued in Ayurvedic Rasayana like *Asparagus racemosus*, *Achillea wilhelmsii*, *Acacia catechu*, *Adhatoda vasica*, *Aloe vera*, *Amorphophallus companulatus*, *Azadirachta indica*, *Bauhinia variegata*, *Bergenia stracheyi*, *Capparis zeylanica*, *Carica papaya*, *Chlorophytum arundinaceum*, *Centella asiatica*, *Curcuma longa*, *Dioscorea alata*, *Enicostemma axillare*, *Justicia spicigera*, *Mangifera indica*, *Madhuca longifolia*, *Ocimum sanctum*, *Ricinus communis*, *Tinospora cordifolia*, *Toxicodendron pubescens*, *Zingiber officinale* etc., are evaluated for their therapeutic potential and have been scientifically investigated with promising results. A number of plant-based principles have been isolated with potential immunomodulatory activity that can explain and justify their use in traditional medicine in the past and can form the basis for further research in the future as well. This work shall hopefully encourage researchers to undertake further work on medicinal plants with potential immunomodulatory activity.

Keywords: Immunomodulatory, Rasayana, Humoral immunity, Cellular immunity, Ayurvedic plants, Haemagglutination, Phagocytosis

INTRODUCTION

Immunomodulation is a very broad term which denotes any changes in the immune response and may involve induction, expression, amplification or inhibition of any part or phase of the immune response. Modulation may be very specific, which is limited to a given antigen/agent or to a non-specific agent with a general effect on immune response. Stimulation of the immune

response is desired for certain people such as immune compromised patient, whereas suppression of the immune response is thought for others, such as transplant recipient, allergic and inflammatory disease patients [1].

Immuno stimulation and immunosuppression both need to be tackled in order to regulate the normal immunological functioning. Hence both immune stimulating agents and immunosuppressing agents have their own standing and search for better agents exerting these activities is becoming the field of major interest all over the world [2].

Every country in the world has lists of herbal remedies for the treatment of diseases and different other unwanted conditions in humans [3]. Ayurveda is one of the traditional systems of medicine practiced in India and Sri Lanka and can be traced back to 6000 B.C [4]. Ayurveda, like other medical systems

*Corresponding Author

Kalliappan Ilango, PhD

Dean, Interdisciplinary School of Indian System of Medicine (ISISM), SRM University, Kattankulathur 603203, Kancheepuram (Dt), Tamil Nadu, India.

Tel: 91-44-27455818; Fax: 91-44-27456702

Email: ilangok67@gmail.com

practised in the world, originated from folk medicine and now holds a commanding position in the China, Germany etc., Ayurvedic medicines are largely based upon herbal and herbo mineral preparations and have specific diagnostic and therapeutic principles [5]. Modulation of immune responses to alleviate the diseases has been of interest for many years and the concept of 'Rasayana' in Ayurveda is based on related principles [4].

Concept of Rasayana

The word Rasayana, a combination of two words (rasa and ayana), refers to nutrition and its transportation throughout the body. Rasayana therapy enhances the qualities of rasa, enriching it with nutrients so one can attain longevity, improved memory and intelligence, freedom from disorder, youthfulness, excellence of hair, complexion and voice, optimum development of physique and sense organs, mastery over phonetics and brilliance. As a dedicated stream of medication for immune promotion, anti-degenerative and rejuvenating health care, the Rasayana therapy of Ayurveda is known to prevent the effects of ageing and improve the quality of life for healthy as well as diseased individuals. Rasayana is helpful to improve immunity and is normally advised during the degenerative phase of life, which starts from around 45 years in both males and females [6].

Chemotherapeutic agents like Cyclosporine, Tacrolimus, Sirolimus, Azathioprine, Cyclophosphamide etc., which are available today mainly possess immunosuppressive activity and most of them are cytotoxic and exerts a variety of side effects like hepatotoxicity [7] gingival hypertrophy, tremor and increased blood pressure, nephrotoxicity [8]. To minimize unwanted side effects, botanicals with immunomodulating activity are getting more popular. Although cytokines like interleukins and interferon's, Levamisole are used as immune stimulants, these are not very effective in the long term both because of their cost and adverse effects like agranulocytosis, hypersensitivity etc., Thus, medicinal plants and their active components as a source of immune modulatory agents are gaining importance. Uses of plant products to enhance the phagocytic ability of macrophages and increase the antibody production by B cells have been well documented by several researchers [9-13].



Source of Photos:

A: <http://avanichef.com/healing-herbs-spices/ingredients/shatavari-asparagus-racemosus/>

B: <http://www.tcfdata.org/?lang=en&page=product-detail&id=62>

C: http://www.indiatradelead.com/product-detail.asp?pr_id=226&ti=Acacia%20Catechu

Ayurvedic plants having Immuno modulating activity

A. *Asparagus racemosus* Willd. (Liliaceae)

The aqueous root extract of the *Asparagus racemosus* having potential immunoadjuvant that also offers direct therapeutic benefits in less morbidity and mortality. It shows significant Immuno protection against intra-cerebral challenge of live *Bordetella pertussis* cells was evaluated based on degree of sickness, paralysis and subsequent death. Reduced mortality accompanied with overall improved health status was observed in treated animals after intra-cerebral challenge of *B. pertussis* indicating development of protective immune response. Fructo-oligosaccharides from the aqueous extract potentiate natural killer cell activity and it could be an important mechanism underpinning the 'Rasayana' properties of this plant [14, 15].

B. *Achillea wilhelmsii* C.Koch (Asteraceae)

The aqueous extract of *Achillea wilhelmsii* exhibited immuno stimulant activity on both humoral and cellular immune functions in mice at a dose of 100mg/kg. It shows significant increase in the delayed type hypersensitivity at a dose of 100mg/kg also shows stimulatory effect on Haemagglutination titre at all doses [16].

C. *Acacia catechu* Wild.var. (Leguminosae)

Aqueous extract of *Acacia catechu* at a dose of 50mg/kg shows significant increase in the neutrophil adhesion to the nylon fibers produced a significant increase in the phagocytic index and a significant protection against cyclophosphamide induced neutropenia indicating its effect on cell mediated immunity. On the other hand, *Acacia catechu* extract produced a significant increase in the serum immunoglobulin levels, increase in the haemagglutination titre values and decreased the mortality ratio in mice, suggesting its effect on the humoral arm of the immune system [17].

A. D. *Adhatoda vasica* Linn (Acanthaceae)

B. Oral administration of methanolic, chloroform and diethyl ether extracts of leaves *Adhatoda vasica* at a dose of 400mg/kg in adult male Wistar rats significantly increased the percentage neutrophil adhesion to nylon fibers and also found to induce delayed type hypersensitivity reaction

by sheep erythrocytes were pharmacologically validated for its immunomodulatory properties in experimental animals. Vinothapooshan et al., suggested that the extracts of this plant positively modulates the immunity of the host [18].

E. *Aloe vera* (L.) Burn. (Liliaceae)

Intraperitoneal administration of *Aloe vera* extract to Swiss albino mice at a dose of 300mg/kg daily for five days significantly increased the total white blood cells count, humoral immune response and plaque-forming cells in the spleen and circulating antibody titre [19].

F. *Amorphophallus campanulatus* Roxb. (Araceae)

Tripathi et al., suggested that the methanolic extract of *A. campanulatus* tuber significantly and dose dependently suppressed the immune system in mice. It exhibits immunomodulatory activity by causing a significant decrease in charcoal clearance, spleen index and delayed-type hypersensitivity response [20].

G. *Azadirachta indica* A. Juss (Meliaceae)

Aqueous extract of the stem bark of *Azadirachta indica* showed strong anticomplementary effects which were dose and time-dependent and most pronounced in the classical complement pathway. Intraperitoneal administration of the neem oil acts as a non-specific immunostimulant and that it selectively activates the cell-mediated immune mechanisms to elicit an enhanced response to subsequent mitogenic or antigenic challenge [21]. Pretreatment of the NIM-76 a volatile fraction from Neem oil

resulted in an increase in polymorphonuclear leucocytes with a concomitant decrease in lymphocyte counts. NIM-76 acts through cell-mediated mechanisms by activating macrophages and lymphocytes [22].

H. *Bauhinia variegata* Linn. (Caesalpiniaceae)

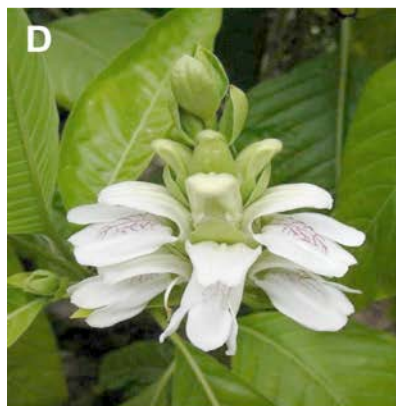
Ethanol extract of the stem bark of *Bauhinia variegata* significantly increases the phagocytic index and percentage neutrophil index at doses of 250 and 500 mg/kg/p.o. It holds a promise as an immunomodulating agent, which acts probably by stimulating both the specific and nonspecific arms of immunity [23].

I. *Bergenia stracheyi* Hook & Thorns (Saxifragaceae)

Oral administration of Bergenin, a C-glycoside of 4-O-methyl gallic acid, isolated from rhizomes of *Bergenia stracheyi* and its O-demethylated derivative norbergenin, prepared from bergenin, are reported to show possible modulation of Th1/Th2 cytokine balance. Flow cytometric study showed that these compounds at doses of 5, 10, 20, 40 and 80 mg/kg per oral dose inhibit the production of proinflammatory Th1 cytokines (IL-2, IFN- α and TNF- α) while as potentiate anti-inflammatory Th2 cytokines (IL-4 and IL-5) in the peripheral blood of adjuvant-induced arthritic balb/c mice. This shows the potential Th1/Th2 cytokine balancing activity of **bergenin** and **its derivative** which is strongly correlated with their anti-arthritic activity [24].

J. *Capparis zeylanica* Linn. (Capparidaceae)

Capparis zeylanica commonly known as Indian caper, is a climbing



Source of Photos:

D: <http://www.herbal-supplement-resource.com/adhatoda-vasica.html>; E: <http://healthandcare.in/what-are-the-benefits-of-aloe-vera/>

F: <http://www.tarladalal.com/glossary-yam-485i>; G: <http://www.treknature.com/gallery/Asia/India/photo164027.htm>

H: http://www.flickriver.com/photos/dinesh_valke/3128061459/; I: http://farm9.staticflickr.com/8152/7325830268_4640367783.jpg

shrub found throughout India and has been used as a rasayan drug in the traditional Ayurvedic system of medicine. Ethanollic and aqueous extracts of leaves on cellular and humoral immune responses to the antigenic challenge by SRBCs and by neutrophil adhesion test, phagocytic activity and cyclophosphamide-induce myelosuppression. Pretreatment of the aqueous extract of leaves of *Capparis zeylanica* shows significant increase in the neutrophil adhesion to nylon fibers. A dose related increase in both primary and secondary antibody titre was observed. *Capparis zeylanica* extracts prevented myelosuppression in mice treated with cyclophosphamide drug [25].

K. *Carica papaya* Linn. (Caricaceae)

Aqueous leaf extract of *Carica papaya* shows significant growth inhibitory activity on tumor cell lines and the production of IL-2 and IL-4 was reduced following the addition of extract, whereas IL-12p40, IL-12p70, IFN- α and TNF- α was enhanced without growth inhibition. Microarray analyses showed that the expression of 23 immuno modulatory genes, classified by gene ontology analysis, was enhanced by the addition of extract. *Carica papaya* leaf extract can mediate a Th1 type shift in human immune system and suggested that the leaf extract may potentially provide the means for the treatment and prevention of selected human diseases such as cancer, various allergic disorders and may also serve as immune adjuvant for vaccine therapy [26].

L. *Chlorophytum arundinaeceum* Baker (Liliaceae)

Oral administration of hydro alcoholic extract of roots of *Chlorophytum arundinaeceum* showed a significant increase in the humoral antibody responses, by increasing the hemagglutinating antibody titre at doses of 100 and 200 mg/

kg/p.o in Swiss albino mice. There was a significant dose dependent increase in the phagocytic index and percentage neutrophil adhesion. It acts probably by stimulating both the specific and nonspecific arms of immunity [27].

M. *Centella asiatica* Linn (Apiaceae)

Methanolic extracts of *Centella asiatica* (0.18% w/v of Asiaticoside) and *Eclipta alba* (1.6% w/v Wedelolactone) at five doses of 100, 200, 300, 400 and 500 mg/kg b.w had significantly increased the phagocytic index and total WBC Count evaluated by using carbon clearance test and cyclophosphamide immunosuppression method [28].

N. *Curcuma longa* Linn (Zingiberaceae)

Administration of 50 mg/kg *Curcuma longa* rhizome in Swiss albino mice protects from the damaging actions CCl₄ on the non-specific host response in the peritoneal macrophages of these CCl₄ intoxicated mice. It has immunotherapeutic properties along with its ability to ameliorate hepatotoxicity [29]. High polarity fraction of the hot water extract exhibited stimulatory effects on human peripheral blood monocytes proliferation. The cytokine productions (TGF- α , TNF- β , GM-CSF, IL-1 α , IL-5, IL-6, IL-8, IL-10, IL-13, etc.) have been modulated by a polysaccharide-enriched fraction. It acts as an adjuvant supplement for cancer patients, whose immune activities were suppressed during chemotherapies [30].

O. *Dioscorea alata* L. cv. Tainong (Dioscoreaceae)

Dioscorin isolated from the tuber of *Dioscorea alata* was evaluated for its immune modulatory activity *in vitro* in the presence of polymyxin B to eliminate lipopolysaccharide contamination. 5-100 μ l of Dioscorin was able to stimulate nitric oxide production



Source of Photos:

J: <http://www.flickr.com/photos/32132262@N03/4611167475/>; K: <http://www.agripinoy.net/medicinal-plant-papaya-carica-papaya.html>

L: <http://www.salenatural.net/impotence-treatment-calipus/>; M: <http://www.spicesmedicinalherbs.com/centella-asiatica.html>

N: <http://www.mgtshop.creamjula.com/index.php/acne-clear-serum.html>; O: <http://www.onlyfoods.net/dioscorea-alata.html>

in RAW264.7 cells. Dioscorin was found to induce IL-6, TNF- α , and IL-1 β production in RAW264.7 cells and human monocytes. The stimulated proliferation index of splenic cells ranged from 1.38 to 1.48 fold of phytohaemagglutinin (PHA) alone for PHA mixed with different concentrations of dioscorin (10, 25, and 50 mg/ml). Yen-Wenn Liu et al., suggested that, the tuber storage protein of yam dioscorin functions as an immunomodulatory substance [31].

P. *Enicostemma axillare* (Lam) A.Raynal. (Gentianaceae)

The methanol extract of *Enicostema axillare* act on both humoral and cell mediated immune functions and decreases the release of pro-inflammatory cytokines in the peritoneal macrophages. Treated with extract appeared to give phagocytic modulation without dose response relationship. Higher concentrations of the extract showed a smaller inhibitory effect and it inhibits the production of NO⁻ significantly in a dose dependent manner. Presence of the compound swertiamarin in crude extract is responsible for the activity [32].

Q. *Justicia spicigera* Schltldl (Acanthaceae)

Ethanol extract of leaves of *Justica spicigera* administrated at doses of 10, 50 and 100 mg/kg i.p. inhibited the tumor growth by 28%, 41% and 53% respectively, in mice bearing human cervical carcinoma cell (HeLa) tumor. *J. spicigera* extract stimulated the phagocytosis of *Saccharomyces cerevisiae* yeasts and the free radicals released by human differentiated

macrophages and also stimulated the proliferation of murine splenocytes and induced natural killer cell activity. [33].

R. *Mangifera indica* Linn (Anacardiaceae)

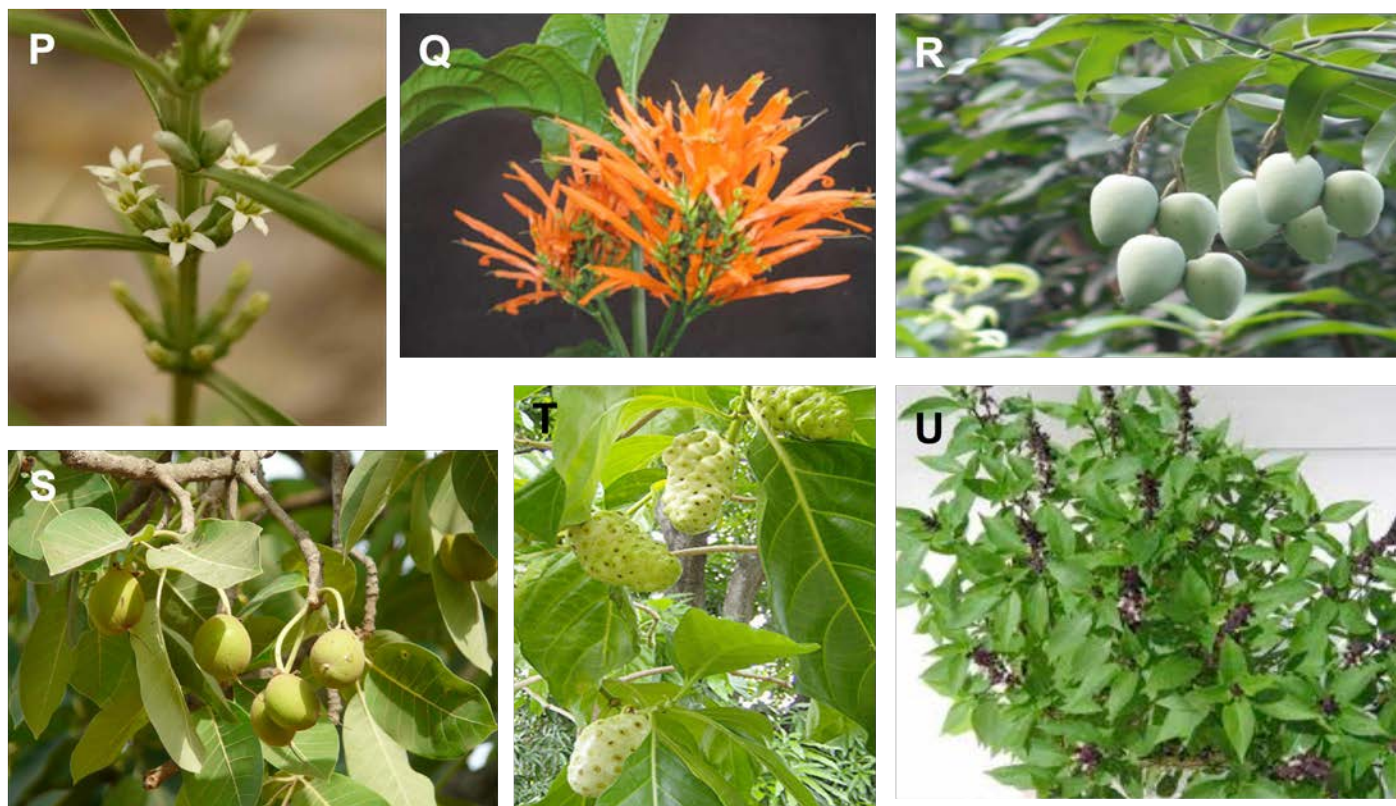
The alcoholic extract of stem bark of *Mangifera indica* containing mangiferin 2.6% has been investigated for its effect on cell mediated and humoral components of the immune system in mice. Administration of extract to the Swiss albino mice at a dose of 50 to 800mg/kg per oral, higher doses 200,400 and 800mg/kg produces significant increase in humoral antibody titre and delayed type hypersensitivity in mice and the extract is having a promising drug with immunostimulant properties [34].

S. *Madhuca longifolia* Var.latifoli (Sapotaceae)

Ethanol extract of the whole plant of *Madhuca latifolia* shows an enhanced humoral immune response on 7th day by 13% as compared to the cyclophosphamide that exhibited 54% humoral immune response, whereas cell mediated immune response was observed with an enhancement in the values (20.27%) when compared with cyclophosphamide (37.63%) [35].

T. *Morinda citrifolia* Linn (Rubiaceae)

The stimulatory effects of the extracts of *M.citrifolia* fruits on important components of the adaptive immune system such as T lymphocytes and B lymphocytes were studied. The effects of the plant extracts on lymphocytes were assessed by *in vitro* MTT



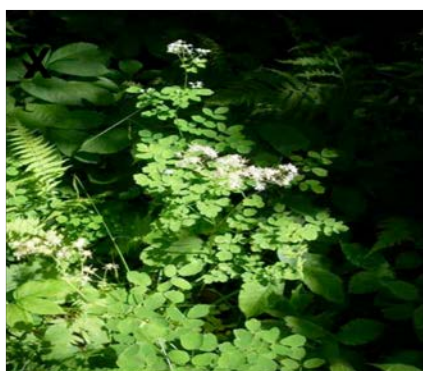
Source of Photos:

P: http://www.zimbabweflora.co.zw/speciesdata/imagedisplay.php?species_id=144700&image_id=2; Q: <http://www.rareflora.com/justiciaspi.html> R: <http://www.mdidea.com/products/herbextract/mangiferin/photogallery.html>; S: http://www.flickr.com/photos/dinesh_valke/3558300625/gallery/Asia/India/photo164027.htm; T: http://en.wikipedia.org/wiki/Morinda_citrifolia; U: http://mbbsstudentcorner.blogspot.in/2011_04_01_archive.html

assay and *in vivo* (Cell mediated immune response) techniques. The MTT study indicated that the hydroalcoholic (0.5 and 1.0mg/mL) and aqueous extracts (0.5 and 1.0mg/mL) significantly ($p < 0.05$) increased *in vitro* splenocyte proliferation to the extent of 43.6, 54.5, 32.7, and 36.4%, respectively. Moreover, the hydroalcoholic (200 mg/kg) and the aqueous (200 mg/kg) extracts significantly ($p < 0.05$) increased the cell-mediated immune response to the extent of 33.52 and 18.56%, respectively [36].

U. *Ocimum sanctum* Linn. (Meliaceae)

Aqueous leaf extract of *Ocimum sanctum* have stimulatory effect on T and B lymphocytes particularly on T-helper subset of lymphocytes by enhancement in IL-2 production. The extract shows the significant enhanced ability of spleen cells to secrete IL-2. It improves overall body constitution by increasing haemoglobin, leukocytosis and lymphocytosis [37]. Several researchers have confirmed repeatedly that *O. sanctum* increases both specific and nonspecific immunity [38, 39, 40]. *Ocimum sanctum* seed oil significantly increased the anti-sheep red blood cells antibody titre value and decreases the percentage of histamine release from peritoneal mast cells of sensitized rats (humoral immune responses) and also decreased the footpad thickness and percentage of leucocyte migration inhibition. It appears to modulate both humoral and cell-mediated immune responsiveness and these immune modulatory effects may be mediated by GABAergic pathways [41]. 300mg capsules of ethanolic extracts of leaves of *O. sanctum* was through a double blind randomized controlled cross-over trial on healthy volunteers shows statistically significant increase in the levels of



Source of Photos:

V: http://dtirp.dtra.mil/CBW/news_archive/toxins.aspx; W: <http://kumarpati.wordpress.com/2010/08/27/guduchi-tinospora-cordifolia/>
X: <http://www.tarlalal.com/glossary-yam-485i>; G: <http://www.treknature.com/gallery/Asia/India/photo164027.htm>
H: http://www.flickrriver.com/photos/dinesh_valke/3128061459/; I: http://farm9.staticflickr.com/8152/7325830268_4640367783.jpg

IFN- γ , IL-4 and percentages of T-helper cells and Natural killer cells were observed after 4 weeks in the extract intervention group in contrast to the placebo group [42].

V. *Ricinus communis* Linn (Euphorbiaceae)

The isolated tannin compounds from the methanolic leaf extract of *Ricinus communis* significantly increased the phagocytic function of human neutrophils and neutrophil chemotactic movement as indicated by the increase in number of cells reaching the lower surface of filter; therefore, the study result revealed that, the extract acts as a chemo attractant which exhibited by increase in the intracellular reduction of Nitroblue tetrazolium dye to formazan by the neutrophils and confirms the intracellular killing property and overall metabolic integrity of phagocytosing neutrophils. Tannins obtained from the extract possess anti-inflammatory and immune modulatory properties [43].

W. *Tinospora cordifolia* (Willd) Miers ex Hook. F & Thomas (Menispermaceae)

Ethylacetate, water fraction and hot water extract of stem of *Tinospora cordifolia* significantly increased the phagocytic function of human neutrophils. Seven compounds of different classes such as alkaloids, phenylpropanoids and sesquiterpenes were isolated from the active fractions out of which two cordifolioside A and syringin were reported to have immune modulatory activity [44, 45, 46]. Remaining compounds showed significant enhancement in phagocytic activity and increase in nitric oxide reactive oxygen species generation at a concentration between 0.1 to 2.5 $\mu\text{g/ml}$ [47]. The immuno modulatory activity of *T. cordifolia* may be attributed to the synergistic effect of groups of compounds present in the particular fraction.

X. *Toxicodendron pubescens* Mill (Anacardiaceae)

10 mg/kg of the powdered crude sample of *T. pubescens* in 0.5% carboxy methyl cellulose shows the stimulation of phagocytosis, Candidacidal activity and chemotaxis of human PMN cells [48].

Y. *Zingiber officinale* Roscoe (Zingiberaceae)

The volatile oil of ginger (0.001 to 10 ng/mL) significantly inhibited T lymphocyte proliferation, decreased the number of the total T lymphocytes and T helper cells in a concentration-dependent manner, but increased the percentage of T suppressor cells to the total T lymphocytes in the mice. In addition, the volatile oil of ginger inhibited IL-1 α secretion by the mice peritoneal macrophages in a concentration-dependent manner. Oral administration of the volatile oil of ginger in the doses of 0.125, 0.25 and 0.5 g/kg body weight dose-

independently weakened the delayed type of hypersensitivity response to 2, 4-Dinitro-1-fluorobenzene in the sensitized mice. Hua-li Zhou et al [49] suggested that it influences both cell-mediated immune response and nonspecific proliferation of T-lymphocyte, and may exert beneficial effects in a number of clinical conditions, such as chronic inflammation and autoimmune diseases.

CONCLUSION

From the above review it should be evident that, there are many medicinal plants which exert immune modulatory activity in experimental models at a particular dose. Different types of screening methods both *in vivo* and *in vitro* have been employed to determine their pharmacological activity. Some medicinal plants may stimulate the immune system, (e.g., *Ocimum sanctum*, *Tinospora cordifolia*) and some may suppress the immune response. Also, various secondary metabolites (e.g., alkaloids, glycosides, saponins, flavonoids, coumarins and sterols) exhibit a wide range of immune modulating activity. This review describes the variety of approaches that have been undertaken to establish the immune modulating activity of Indian herbal plants.

REFERENCES

1. Sell S: **Immunomodulation. In Immunology, Immunopathology and Immunity**, Elsevier Science Publishing Company, Inc. New York, 1987, 655-683.
2. Patwardhan B, Kalbag D, Patki PS, Nagsampagi BA: **Search of Immune Modulatory Agents: A Review. Indian Drugs** 1990, 28: 348-358.
3. Satoskar R.S. and Bhandarkar S.D: **Pharmacology and Pharmacotherapeutics**, Part I, 8th ed. Popular Prakashan, Bombay, India. 1983.
4. Charak Samhita, **Chikitsa Sthana** 1:4, Shree Gulabkunverba Ayurvedic Society. Jamnagar, India, 1949.
5. Patwardhan B and Hooper M: **Ayurveda and Future Drug Development. International J Altern Complem Med** 1992, 10: 9-10.
6. Rita rezzani: **Cyclosporine A and Adverse Effects on Organs: Histochemical Studies. Prog Histochem Cyto** 2004, 39: 85-128.
7. Chulet R and Pradhan P: **A Review on Rasayana. Phcog Rev** 2010, 3: 229e34.
8. Tolba R.H, Wrigge B, Machein U, Minor T, Kalff JC, Hirner A, Wolff M : **Conversion to Neoral for Tacrolimus-Related Adverse Effects in Liver Transplant Recipients and Improvement in Quality of Life. Transplant** 2001, 33: 3446-3447.
9. Chopra R.N, Nayar S.L and Chopra I.C: **Glossary of Indian Medicinal Plants**. First ed. CSIR, New Delhi; 2006.
10. Chopra R.N, Chopra L.C, Handa K.D, and Kapur L.D: **Indigenous Drugs of India**, Second ed. M/S Dhar, V.N. and Sons, Kolkata; 1982.
11. Nadkarni K.M, Nadkarni, A.K: **Indian Materia Medica**, 3rd ed. M/S Popular Prakashan Pvt. Ltd, Mumbai; 2009.
12. Kirtikar K.R, Basu, B.D, **Indian Medicinal Plants**, 2nd ed. Periodical Expert book agency, New Delhi, India; 2012.
13. Zhao T.F, Wang X, Rimando A.M and Che C: **Folkloric Medicinal Plants: *Tinospora sagittata* var. *cravaniensis* and *Mahonia bealei*. Planta Med** 1991, 57: 505-510.
14. Manish Gautam, Santanu Saha, Sarang Bani, Kaul A, Sanjay Mishra, Dada Patil, Satti N.K., Suri K.A., Sunil Gairola, Suresh K et al.: **Immuno adjuvant potential of *Asparagus racemosus* aqueous extract in experimental system. J Ethnopharmacol** 2004, 91:251-255.
15. Mayank Thakur, Connellan Paul A, Myrna A.D, Carol Moris A, Werner Praznik, Loeppert, Dixit et al: **Characterization and in vitro immune modulatory screening of fructo-oligosaccharides of *Asparagus racemosus* Willd. Int J Biol Macromol** 2012, 50:77- 81.
16. Fariba sharififar, Shirin pournourmahammadi and Moslem Arabnejad: **Immunomodulatory activity of aqueous extract of *Achillea wilhelmsii* C.Koch in mice. Indian J Exp Biol** 2009, 47: 668-671.
17. Syed Ismail and Mohammed Asad: **Immuno modulatory activity of *Acacia catechu*. Indian J Physiol Pharmacol** 2009, 53:25-33.
18. Vinothapooshan G and Sundar K: **Immuno modulatory activity of various extracts of *Adhatoda vasica* Linn. in experimental rats. African J Pharm Pharmacol** 2011, 5:306-310.
19. Jyotsana madan, Arun kumar Sharma, Nazma Inamdar, Harwinder Singh Rao, Ramnik Singh: **Immuno modulatory properties of *Aloe vera* gel in mice. Int J Green Pharm** 2008, 2:152-154.
20. Tripathi AS, Chitra V, Sheikh NW, Mohale DS and Dewani AP: **Immuno modulatory Activity of the Methanol Extract of *Amorphophallus campanulatus* (Araceae) Tuber. Trop J Pharm Res** 2010, 9: 451- 454.
21. Shakti N. Upadhyay, Suman Dhawan, Sanjay Garg, GP. Talwar: **Immuno modulatory effects of neem (*Azadirachta indica*) oil, Ann Limnol** 1992, 14:1187-1193.
22. Sai Ram M, Sharma S.K, Ilavazhagan G, Devendra Kumar, Selvamurthy W : **Immuno modulatory effects of NIM-76, a volatile fraction from Neem oil. J Ethnopharmacol** 1997, 55:133-139.
23. Ghasias M.M, Shaikh S.A, Deshpande A.D: **Evaluation of the immuno modulatory activity of the ethanolic of the stem bark of *Bauhinia variegata* Linn. Int J Green Pharm** 2006, 3:70-74.
24. Nighat Nazir, Surrinder Koul, Mushtaq Quireshi, Sachin C Taneja, Sheikh F Ahmed, Sarang Bani, Ghulam N Qazi: **Immuno modulatory effect of bergenin and norbergenin against adjuvant-induced arthritis - A flow cytometric study. J Ethnopharmacol** 2007, 112:401-405.
25. Ghule BV, Murugananthan G, Nakhat PD, Yeole PG: **Immunostimulant effects of *Capparis zeylanica* Linn. Leaves. J Ethanopharmacol** 2006, 108:311-315
26. Noriko Otuski, Nam H Dang, Emi Kumagai, Akira Kondo, Satoshi Iwata, Chikao Morimoto: **Aqueous extract of *Carica papaya* leaves exhibits anti-tumor activity and immune modulatory effects. J Ethnopharmacol** 2010, 127:760-767.

27. Ananth kumar K, Chitra, Leela Krishna V, Soujanya J: **Evaluation of immune modulatory effect of hydroalcoholic root extract of *Chlorophytum arundinaeceum baker* in mice.** *Int J Pharm Pharm Sci* 2011, 3:355-359.
28. Jayathirtha M.G, Mishra S.H: **Preliminary immune modulatory activities of methanol extracts of *Eclipta alba* and *Centella asiatica*.** *Phytomedicine* 2004, 11: 361-365.
29. Mahuya Sengupta, Gauri Dutta Sharma, Biswajit Chakraborty: **Hepatoprotective and immune modulatory properties of aqueous extract of *Curcuma longa* in carbon tetra chloride intoxicated Swiss albino mice.** *Asian Pac J Trop Med* 2011, 3: 193-199.
30. Grace G.L. Yu, Ben C.L.Chan, Clara B.S.Lau.: **Immuno stimulatory activities of polysaccharide extract isolated from *Curcuma longa*.** *Int J Biol Macromol* 2010, 47:342-347.
31. Yen-Wenn Liu, Shang H.F, Wang C K, Hsu F.L, Hou W.C : **Immuno modulatory activity of dioscorin, the storage protein of yam (*Dioscorea alata* cv. Tainong No. 1) tuber.** *Food Chem Toxicol* 2007, 45:2312-2318.
32. Saravanan S, Prakash Babu N, Pandikumar P, Karunai Rai M, Gabriel Paulraj M, Ignacimuthu S: **Immuno modulatory potential of *Enicostema axillare* (Lam.) A. Raynal, a traditional medicinal plant.** *J Ethnopharmacol* 2012, 140:239- 246.
33. Angel Josabad Alonso-Castro, Ortiz-Sanchez E, Dominquez F, Arana-Argaez V, Juarez-Vazquez Mdel C, Chavez M, Carranza-Alvarez C, Gaspar-Ramirez O, Espinosa-Reyes G, Lopez-Toledo G, Ortiz-Ardrade R, Garcia Carranca A: **Antitumor and immune modulatory effects of *Justicia spicigera* Schltdl (Acanthaceae).** *J Ethnopharmacol* 2012, 141: 888- 894.
34. Neelam Makare, Subhash Bodhankar, Vinod Rangari: **Immuno modulatory activity of alcoholic extract of *Mangifera indica* L. in mice.** *J Ethnopharmacol* 2001, 78:133-137.
35. Reena Purohit, Santosh Kr Singh, Asima Shaban, Rajesh nautiyal, Purohi M.C, Satish K.Verma: **Immunomodulatory activity of *Madhuca longifolia*.** *Der Pharmacia Sinica*, 2012, 3:153-155
36. Nayak S, Mengi S: **Immunostimulant activity of noni (*Morinda citrifolia*) on T and B lymphocytes.** *Pharma Biol* 2010, 48:724-731.
37. Anjana Goel, Dilip Kumar Singh, Sandeep Kumar, Ashok Kumar Bhatia: **Immuno modulating property of *Ocimum sanctum* by regulating the IL-2 production and its mRNA expression using rats splenocytes.** *Asian Pac J Trop Med* 2010, 3: 8-12.
38. Godhwani s, Godhwani JL, Vyas DS: ***Ocimum sanctum*: an experimental study evaluating its anti-inflammatory, analgesic and antipyretic activity in animals.** *J Ethnopharmacol* 1987, 21:153-163.
39. Godhwani S, Godhwani JL, Vyas DS: ***Ocimum sanctum*: a preliminary study evaluating its immunoregulatory profile in albino rats.** *J Ethnopharmacol* 1988, 24:193-198.
40. Mediratta PK, Dewan V, Bhattacharya SK, Gupta VS, Maiti PC, Sen P: **Effect of *Ocimum sanctum* Linn. On humoral immune response.** *Indian J Med Res* 1988, 87: 384-386
41. Mediratta P.K, Sharma K.K, Surender Singh: **Evaluation of immune modulatory potential of *Ocimum sanctum* seed oil and its possible mechanism of action.** *J Ethnopharmacol* 2002, 80:15-20.
42. Shankar mondal, Varma S, Bamola VD, Naik SN, Mirdha BR, Padhi MM, Mehta N, Mahapatra SC: **Double-blinded randomized controlled trial for immune modulatory effects of Tulsi (*Ocimum sanctum* Linn.) leaf extract on healthy volunteer.** *J Ethnopharmacol* 2011, 136: 452- 456.
43. Aravind Kumar, Vaishali Singh, Sourav Ghosh: **An experimental evaluation of *In vitro* immune modulatory activity of isolated compounds of *Ricinus communis* on human neutrophils.** *Int J Green Pharm* 2011, 5: 201-204.
44. Maurya R, Wazir V, Kapil A, Kapil R.S: **Cordifolisides A and B, two new phenylpropene disaccharides from *Tinospora cordifolia* possessing immune stimulant activity.** *Nat Prod Lett* 1996, 8: 7-10.
45. Kapil A, Sharma S: **Immuno potentiating compounds from *Tinospora cordifolia*.** *J Ethnopharmacol* 1997, 58: 89-95.
46. Cho J.Y, Nam K.H, Kim A.R, Park J, Yoo E.S, Baik K.U, Yu Y.H, Park M.H : ***In vitro* and *In vivo* immune modulatory effects of syringin.** *J Pharm Pharmacol* 2001, 53: 1287-1294.
47. Upendra Sharma Bala M, Kumar N, Singh B, Munshi RK, Bhalerao S: **Immuno modulatory active compounds from *Tinospora cordifolia*.** *J Ethnopharmacol* 2012, 141: 918- 926.
48. Patil CR, Salunkhe PS, Gaushal MH, Gadekar AR, Agrawal AM, Surana SJ : **Immuno modulatory activity of *Toxicodendron pubescens* in experimental models.** *Homeopathy* 2009, 98:154-159.
49. Hua-li Zhou, Yang-mei Deng, Qiang-min Xie: **The modulatory effects of the volatile oil of ginger on the cellular immune response *In vitro* and *In vivo* in mice.** *J Ethnopharmacol* 2006,105: 301-305.

Note: Vedic Research International, Vedic Research Inc is not responsible for any data in the present article including, but not limited to, writeup, figures and tables. If you have any questions, directly contact authors.

Visit us @ www.vedicjournals.com :

DOI: <http://dx.doi.org/10.14259/pm.vii2.39>